

REVIEW

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Beyond the anti-PD-1/PD-L1 era: promising role of the BTLA/HVEM axis as a future target for cancer immunotherapy

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Abstract

Recent introduction of monoclonal antibodies targeting immune checkpoints to harness antitumor immunity has revolutionized the cancer treatment landscape. The therapeutic success of immune checkpoint blockade (ICB)-based therapies mainly relies on PD-1/PD-L1 and CTLA-4 blockade. However, the limited overall responses and lack of reliable predictive biomarkers of patient's response are major pitfalls limiting immunotherapy success. Hence, this reflects the compelling need of unveiling novel targets for immunotherapy that allow to expand the spectrum of ICB-based strategies to achieve optimal therapeutic efficacy and benefit for cancer patients. This review thoroughly dissects current molecular and functional knowledge of BTLA/HVEM axis and the future perspectives to become a target for cancer immunotherapy. BTLA/HVEM dysregulation is commonly found and linked to poor prognosis in solid and hematological malignancies. Moreover, circulating BTLA has been revealed as a blood-based predictive biomarker of immunotherapy response in various cancers. On this basis, BTLA/HVEM axis emerges as a novel promising target for cancer immunotherapy. This prompted rapid development and clinical testing of the anti-BTLA blocking antibody Tifcemalimab/icatolimab as the first BTLA-targeted therapy in various ongoing phase I clinical trials with encouraging results on preliminary efficacy and safety profile as monotherapy and combined with other anti-PD-1/PD-L1 therapies. Nevertheless, it is anticipated that the intricate signaling network constituted by BTLA/HVEM/CD160/LIGHT involved in immune response regulation, tumor development and tumor microenvironment could limit therapeutic success. Therefore, in-depth functional characterization in different cancer settings is highly recommended for adequate design and implementation of BTLA-targeted therapies to guarantee the best clinical outcomes to benefit cancer patients.

Keywords Immunotherapy, BTLA, HVEM, Checkpoint blockade, T cell, NK cell, CD160, LIGHT, Icatolimab, Tifcemalimab

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Introduction

The immune response is tightly regulated by a complex network of co-stimulatory and co-inhibitory signals mediated by immune checkpoints, which represents an essential mechanism of immunological homeostasis. In the context of cancer, dysregulation of immune checkpoints promotes immune evasion and prevents tumor elimination by cytotoxic lymphocytes, thereby favoring tumor development and progression [1].

The introduction of immune checkpoint blockade (ICB)-based therapies has revolutionized the landscape of cancer treatment within the last few years. The emergence of immunotherapy was supported by the outstanding response rates achieved in several tumors. Moreover, ICB has become a cornerstone for the treatment of tumors classically linked to poor prognosis, such as melanoma, renal carcinoma, or lung cancer, even evolving/turning into first-line therapies in certain cases [1, 2]. Nonetheless, despite these initial encouraging results, response rates to PD-1 and CTLA-4 blocking antibodies do not exceed approximately 20–30%. Furthermore, various tumors, including pancreatic adenocarcinoma or glioblastoma, exhibit complete resistance to ICB-based therapies, thus supporting the urge of finding new immune checkpoints for cancer immunotherapy [1, 3]. To expand the portfolio of targets with emerging checkpoints may also have the potential to benefit a wider range of patients. Within the last years, a plethora of new co-inhibitory molecules has been explored and several of them are currently under pre-clinical and clinical development, including LAG-3, NKG2A or ILT2 [4–8].

This review focuses on comprehensively examining current knowledge on the immunobiology of the inhibitory immune checkpoint BTLA (B- and T-lymphocyte attenuator) and its binding partner HVEM (Herpesvirus entry mediator), as well as currently available evidence supporting the clinical application of the BTLA/HVEM axis as a future target for cancer immunotherapy.

BTLA is an immunoinhibitory receptor

The inhibitory immune checkpoint BTLA/CD272 belongs to the CD28 immunoglobulin superfamily. This receptor recognizes and binds to HVEM/CD270, a member of the TNF (tumor necrosis factor) receptor family, portraying the first described interaction or crosstalk between both superfamilies. BTLA is broadly expressed in the immune system, mainly on T and B lymphocytes, macrophages and dendritic cells and, at low levels, on NK cells [9, 10]. Likewise, HVEM is expressed on T and B lymphocytes, as well as NK cells, dendritic and myeloid cells, thus suggesting a complex heterotypic interaction between the different immune cell subsets that express both molecules [10]. Regarding T lymphocytes, BTLA

expression arises during the positive selection that takes place in the thymus, being higher in CD4+ than in CD8+ T lymphocytes and, unlike PD-1 or CTLA-4, with no expression reported on regulatory T cells (Tregs) [11]. BTLA expression can be detected at low levels in the bone marrow during the pro-B and pre-B phase, whereas it is constitutively expressed on naïve B lymphocytes [12].

BTLA signaling negatively regulates immune responses through recruitment of phosphatases 1 and 2 with Src homologous domain (SHP-1 and SHP-2) mediated via two immunoreceptor tyrosine-based inhibitory (ITIM) motifs [13–15]. Noteworthy, BTLA harbors a third domain in its cytoplasmic tail that contains a GRB-2 recognition motif. Recruitment of GRB-2 and the p85 subunit of phosphatidylinositol 3-kinase (PI3K) leads to protein kinase B (PKB or AKT) activation, thus providing pro-survival signaling and bringing forward a dual role of this molecule in the regulation of the immune response (Fig. 1) [15]. Importantly, HVEM functions as a bidirectional molecular switch between activating (CD160, LIGHT, and lymphotoxin LT- α) and inhibitory (BTLA) pathways, depending on the interacting receptor used. Upon binding, HVEM provides pro-survival and proliferative signals through activation of nuclear transcription factor κ B (NF- κ B) and AKT transcriptional pathways, whereas BTLA attenuates T cell-mediated responses [10, 16–18].

The first evidence for the inhibitory function of BTLA was demonstrated in mice deficient for this receptor, which showed an augmented risk of developing autoimmune diseases, such as encephalomyelitis and whose T lymphocytes displayed increased proliferative capacity as well [12, 19, 20]. In line with these *in vivo* observations, *in vitro* studies revealed that BTLA deletion significantly enhanced T cell proliferation upon stimulation with anti-CD3 monoclonal antibodies (mAbs) or peptide-loaded antigen-presenting cells (APCs), while this effect was not observed in the BTLA wild-type (WT) counterparts [12, 20]. Interestingly, BTLA^{-/-} CD8+ T cells showed greater proliferation, despite wild-type CD4+ T lymphocytes exhibited higher BTLA expression [21]. Moreover, larger numbers of memory CD8+ T lymphocytes were detected in BTLA or HVEM KO mice compared to the corresponding WT mice, thus suggesting that blockage of the inhibitory effect of BTLA could increase immune responses to antigenic stimuli [21]. Consistently, treatment of T cells with an agonist anti-BTLA antibody or HVEM-Ig fusion protein led to reduced proliferation and production of cytokines such as IL-2 or IL-4 [22, 23].

Despite previous observations that classified BTLA as an inhibitory immune checkpoint, studies using BTLA^{-/-} graft-versus-host disease (GvHD) mouse models demonstrated that it also plays an important role in

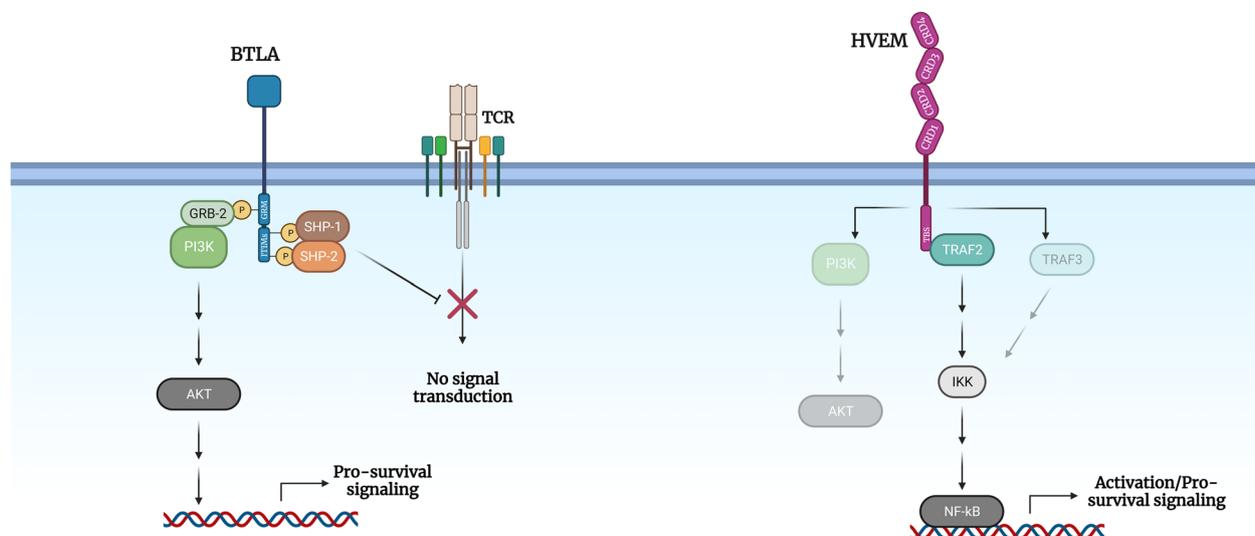


Fig. 1 HVEM binding network regulates immune homeostasis. Activation of BTLA signaling leads to the recruitment of SHP-1/2 and negatively regulates T cell responses, whereas GRB-2 promotes survival through the AKT pathway. HVEM-mediated NF- κ B signaling leads to enhanced activation status, inflammation, and survival

favoring T lymphocyte survival. Donor BTLA^{-/-} T lymphocytes exhibited potent alloreactivity compared to WT during the first week of GvHD response, while being unable to maintain the inflammatory response over time [24]. The significant reduction in BTLA^{-/-} T cell count observed after one week could be related to negative signaling mediated by BTLA-HVEM binding as well as the loss of pro-survival signals via the Grb-2 binding domain of BTLA [15, 24]. In agreement, anti-CD3/CD28 stimulated T cells from BTLA KO mice exhibited enhanced proliferation rates compared to WT mice upon treatment with BTLA-Fc fusion protein by HVEM signaling activation [25]. However, it should be noted that no changes in the number of divisions per cell were found, rather a higher number of T cells were able to proliferate. Therefore, BTLA-Fc-mediated HVEM activation induces pro-survival signaling, thus proposing a protective role for BTLA against cell death.

Currently, little is known about the part that BTLA plays on NK cells. Still, upregulation of BTLA on this immune subset has been observed to compete with CD160, an activating receptor with HVEM-binding ability, dampening NK cell-mediated cytotoxicity and possibly impairing immunosurveillance [26].

HVEM binds to multiple ligands beyond BTLA

HVEM was initially described as the attachment and entry site for herpes virus through binding to the type I and type II gD proteins of the virus [27]. Unlike BTLA, which has an expression pattern mainly restricted to immune cells (both lymphoid and myeloid), HVEM

exhibits a broader expression detected in hematopoietic, epithelial and endothelial cells and neurons [13]. This receptor interacts with members of the TNF superfamily lymphotoxin- α (LT- α) and LIGHT, as well as with CD160 and BTLA, which belong to the immunoglobulin superfamily [28]. In addition, SALM5 has been recently described as a novel binding partner for HVEM and their interaction modulates neuroinflammation by inhibiting the myeloid-related inflammatory response in murine models of multiple sclerosis [29].

HVEM binding to its different ligands is mediated by two extracellular topographical regions constituted by four cysteine-rich domains (CRD), known as CRD1/CRD2 (binding site for BTLA and CD160) and CRD2/CRD3 (binding site for LIGHT and LT α) [14, 30–32]. Due to this structural organization, HVEM acts as a bidirectional switch, acting as ligand for distinct co-stimulatory and co-inhibitory molecules, but also being able to activate its own signal transduction depending on the interacting receptor used. This intricate network is crucial for the homeostatic maintenance of immune responses [31].

Signaling through HVEM is mediated by a TNFR-associated factor 2 (TRAF2) binding site located in its cytoplasmic tail. This initial TRAF2 recruitment ultimately translates into NF- κ B activation, which controls the transcription of genes that promote activation, inflammation and survival [14]. Despite the disparate location of LIGHT, LT- α , CD160, and BTLA binding sites in the extracellular domain of HVEM, all share the ability to activate the canonical NF- κ B pathway [12, 33, 34]. In addition, HVEM stimulation may lead to activation

of non-canonical NF- κ B pathway through recruitment of TRAF3, although this mechanism currently remains unclear [13]. Similarly, HVEM-mediated signaling has been linked to PI3K and AKT signaling pathway in T cells, although the exact mechanism needs to be fully elucidated [35–37].

The TNF superfamily

The HVEM ligands, LIGHT and LT- α , belonging to the TNF superfamily, can be located on the cell surface, or found as soluble forms. While LIGHT has a transmembrane and a soluble form produced by alternative mRNA splicing or proteolytic cleavage of its extracellular domain, LT- α is exclusively assembled as a soluble protein. LT- α homotrimers engage HVEM with low affinity, compared to TNFR1 and 2 receptors. Notoriously, LT- α can also be found on the outer cell surface, throughout its interaction with LT β , which leads to the formation of a heterotrimer. This complex exhibits lower affinity for HVEM, favoring the interaction of the heterotrimer with LT β receptor [38, 39].

LIGHT is a cell-surface homotrimer expressed at low levels on hematopoietic cells, including B and T lymphocytes, as well as dendritic and NK cells, although altered expression has been detected upon activation [40]. Indeed, naïve T cell activation leads to enhanced surface LIGHT expression, whereas, in turn, HVEM levels are decreased [41]. By contrast, although high LIGHT levels and low HVEM are expressed on the surface of dendritic cells at basal state, dendritic cell activation prompts the loss of surface LIGHT and promotes HVEM expression [42]. Altogether, these observations suggest an interaction between dendritic cells and T lymphocytes through the LIGHT/HVEM axis [40–42]. In agreement, several works have reported that LIGHT expression on T lymphocytes and dendritic cells co-stimulates T cell function and boosts cytokine production and proliferation of this immune subset [42–45]. In addition, both soluble and membrane-bound activated T cell-derived LIGHT induce dendritic cell maturation and cytokine production upon HVEM binding [46]. BTLA has been proposed to instruct Treg differentiation in dendritic cells. BTLA+DEC205+CD8+ dendritic cells were revealed as the exclusive mediators of CD5 upregulation in T cells, thereby inducing a "tolerizing" effect. By contrast, CD11c+ dendritic cells lacking BTLA failed to increase CD5 expression, leaving immune responses unchanged. BTLA and HVEM functions enable dendritic cells to connect T cell responsiveness with tolerogenic antigens, influencing CD5-dependent Treg cell induction for peripheral T cell tolerance. The absence of these functions impedes

tolerance induction, thus emphasizing the joint role of BTLA and CD5 in adjusting T cell responses via tolerogenic dendritic cells [47].

The immunoglobulin superfamily

As mentioned above, CD160 binds and activates HVEM through the CRD1 domain [25]. However, this receptor also interacts with MHC class I molecules, both classical (HLA-A, -B and -C) and non-classical (HLA-E, -F and -G) with low affinity [48, 49]. CD160 was initially identified as a glycosylphosphatidylinositol (GPI)-anchored protein displayed on the NK cell membrane. Nonetheless, a transmembrane form derived from alternative mRNA splicing and a soluble form (by the action of metalloproteases) have also been described [50]. Contrarily to HVEM or BTLA, CD160 presents a more restricted expression pattern; the highest levels are detected on CD56dim CD16+ NK cells, although its expression can also be found in $\gamma\delta$ T lymphocytes, NKT cells and activated CD4+ and CD8+ T lymphocytes [51–54].

CD160 exhibits a dual immunomodulatory role. On the one hand, the transmembrane form of CD160 exerts an activating role in NK cells, where treatment with an anti-CD160 agonistic antibody led to an increase of the degranulation marker CD107a, as well as recruitment of the CD107a kinase, Src-family p56 (lck) and activation of the Erk1/2 pathway [50]. In agreement, enhanced NK cell-mediated cytotoxicity via CD160 activation was observed upon transduction of the K562 cell line (characterized by the lack of HLA-I expression) with *TNFRSF14* gene encoding HVEM [26]. In addition, CD160 binding to HLA-C induces activation of CD8+ T lymphocytes and NK cells, as well as an increase in the cytotoxic capability and cytokine production of the latter [48, 49, 55, 56]. Whether CD160 actually plays its part as a co-stimulatory molecule remains controversial, since certain studies have suggested an inhibitory activity for this immune checkpoint. Activation of CD4+ T lymphocytes translates into augmented levels of surface CD160, thus competing with LIGHT for HVEM engagement on the surface of APCs [51]. Additionally, in vitro IL-15-mediated NK cell activation favors cleavage of surface CD160 in a metalloproteases-dependent manner, dampening cytotoxic activity of NK cells and CD8+ T lymphocytes by its binding to HLA-I molecules [57].

The enigmatic activity of CD160 in immune function may indicate that this receptor could act as a molecular switch in some cell types. In this scenario, the extracellular domain of the protein involved in the interaction should define whether negative or positive signals are transduced [40].

Cis and trans interactions modulate the activity of the BTLA/HVEM axis

Significant efforts have been put into deciphering the complex network established by HVEM. Signaling mediated by HVEM and its ligands depends on three parameters that define the immunobiology of this axis: the ligand-receptor pair involved, whether they are in a soluble or membrane-bound form, and the *cis* or *trans* nature of the interaction (Fig. 2).

As already discussed, immune cells co-express HVEM and BTLA on their surface. In a resting immune system, a *cis* interaction -within the same cell- occurs, leading to the formation of HVEM-BTLA heterodimers. This binding suppresses HVEM-mediated *trans* activating signaling (via NF-κB), therefore maintaining this receptor in a resting state [9, 25, 30, 58–60]. As a consequence of immune activation, the surface levels of another HVEM partner, LIGHT, are transiently increased, a regulatory mechanism that prevents an exacerbated response in a steady state, since LIGHT displays a higher avidity for HVEM as compared to BTLA. Further, upon B and T cell activation, there is also a temporary decrease in HVEM levels and a transitory increase in BTLA expression, causing a dissociation of the *cis* complex and allowing BTLA-HVEM *trans* interactions. This strategy ultimately facilitates LIGHT and HVEM binding, and the

consequent activation of the immune response [10, 30]. Although membrane-bound LIGHT can also interact with the HVEM-BTLA heterodimer, it does not induce (or induces at low levels) HVEM signaling cascade. Paradoxically, LIGHT in its soluble form facilitates *cis* interaction stabilization, whereas BTLA or HVEM-Fc fusion proteins cannot even bind to the heterodimeric complex [14]. Despite LIGHT participation in this system, BTLA plays a predominant part in T and B lymphocytes, since it is estimated that the *cis* HVEM-BTLA complex represents nearly 80% of the membrane-expressed proteins [30]. Constitutive HVEM-mediated signaling is detected when co-expressed with LIGHT or CD160, whereas little to no NF-κB pathway activation is achieved upon BTLA-HVEM co-expression, supporting a prevalent BTLA-mediated inhibitory signaling within the *cis* complex. Moreover, BTLA activity has been observed even in *cis* BTLA-HVEM heterodimers. Despite *cis*-heterodimeric complexes prevent *trans* HVEM interactions, recent reports have demonstrated that this binding does not limit BTLA inhibitory pathway [9].

In line with this, BTLA antagonistic antibodies disrupt the *cis* complex, thereby promoting a synergistic effect on HVEM activation via LIGHT [61, 62]. This result suggests that disruption of this complex is mandatory for HVEM activation mediated by the membrane form of

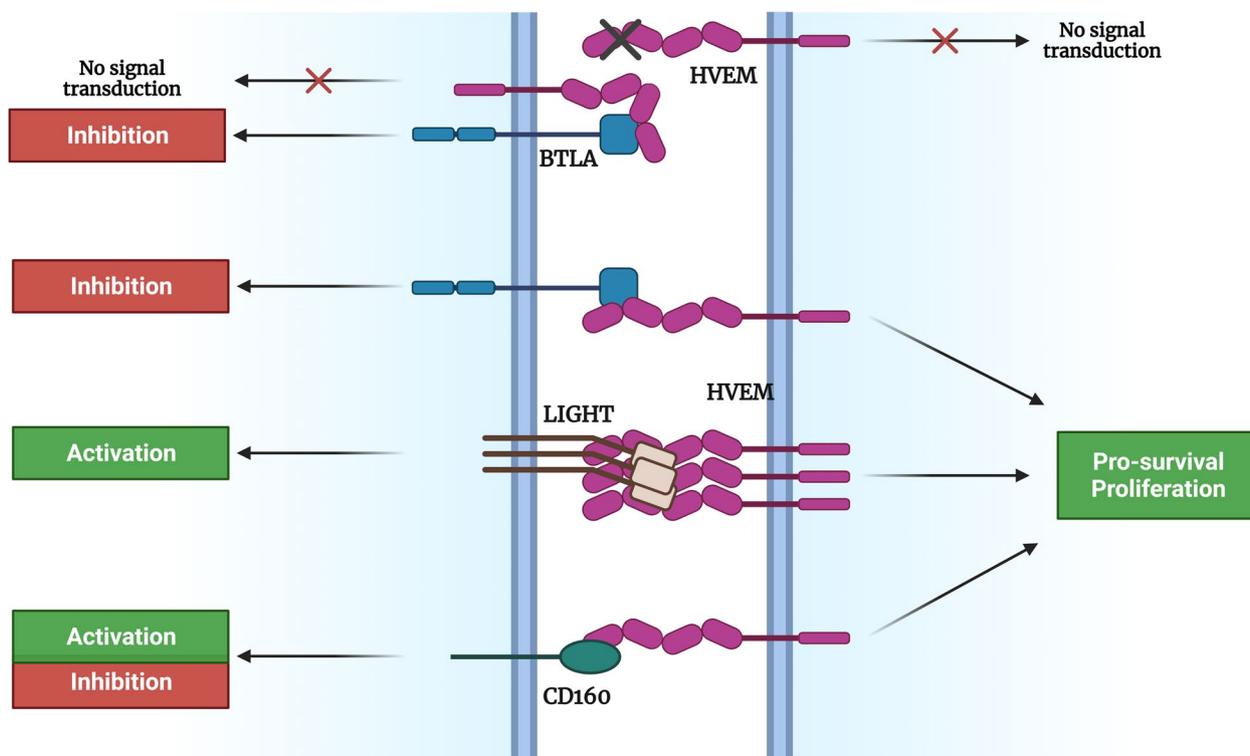


Fig. 2 Signal transduction through HVEM and its binding partners relies on the nature of its interactions (*cis* or *trans*)

LIGHT during the immune response [28]. Nevertheless, Claire Battin et al. have recently outlined the potential benefits from targeting HVEM instead of BTLA. Thus, the use of an agonistic anti-HVEM mAb overcomes BTLA *cis*-mediated inhibition and promotes T cell proliferation, activation and cytokine production, suggesting that it may act as both a stimulating and an immune checkpoint-blocking antibody [9]. Still, the therapeutic advantages of targeting BTLA or HVEM deserve further consideration in the context of cancer or autoimmune diseases, where this axis is often dysregulated.

The BTLA/HVEM axis in solid tumors

T cell exhaustion is a critical phenomenon within the context of immuno-oncology, where the immune system's frontline defenders, including T lymphocytes and NK cells, become progressively dysfunctional and lose their ability to effectively target and eliminate cancer cells. This state of exhaustion arises as a result of chronic antigen exposure and persistent immune stimulation, among others, often leading to altered expression of immune checkpoints in the tumor microenvironment. On this basis, dysregulation of BTLA/HVEM axis has been proposed to play a role in T cell dysfunction using HIV-specific T cells as a model for exhaustion. Antibodies targeting BTLA were found to enhance CD8+ T cell proliferation and cytokine production in response to HIV-1 antigens. In addition, co-targeting PD-1 and BTLA was found to be particularly effective in enhancing responses of exhausted human T cells [63].

In this context, ICB is considered a major breakthrough in the fight against cancer due to its capacity to revert immune exhaustion, and reinvigorate antitumor responses. Noteworthy, ICB is considered a major breakthrough in the fight against cancer. Noteworthy, ICB-based therapies with mAb targeting CTLA-4 or PD-1/PD-L1 promote long-lasting or even curative responses in approximately 20–30% of patients [64]. Yet, a significant percentage of patients do not respond to these immunotherapies, bringing to light the urge to investigate novel potential targets among the broad spectrum of inhibitory receptors that regulate the immune response. In the context of cancer, the immunoregulatory function of HVEM and its ligands, especially BTLA due to its inhibitory nature, is altered, hence contributing to the dysregulation of antitumor immunity (Fig. 3).

Dysregulation of the BTLA/HVEM axis has been described in a plethora of solid tumors and hematological malignancies. In hepatocellular carcinoma, increased percentage of tumor-infiltrating PD-1+ BTLA+ CD4+ T lymphocytes was observed when compared to those from tumor-free regions from the same patient, pointing out a role of the tumor

microenvironment (TME) in this dysregulation [17]. Similarly, elevated surface levels of BTLA on infiltrating CD4+ and CD8+ T lymphocytes in bladder cancer were correlated with poor outcomes [65]. Likewise, patients with non-small cell lung cancer (NSCLC) negative for BTLA and PD-L1 expression exhibited a better prognosis [66]. Moreover, HVEM was found to be overexpressed in 18.6% of NSCLC patients, mainly those with lymph node metastasis and advanced stage [67]. These results are consistent with those obtained in a study with 136 patients with gastric cancer, where high BTLA and HVEM levels were associated with lymph node metastases and diminished overall survival [68]. *BTLA* gene expression in whole blood was augmented in patients with colorectal cancer compared to healthy individuals, which in turn was correlated with the expression of other checkpoints such as LAG-3 or PD-1, as well as lower overall survival [69]. On the other hand, *in silico* analysis of HVEM levels in endometrial cancer unveiled that low mRNA expression was associated with poor prognosis. Besides, *in vitro TNFRSF14* KO showed that this association with decreased survival could be related, at least in part, to enhanced migration and promotion of epithelial-mesenchymal transition (EMT) [70]. These results contrasted with those obtained for melanoma. Using immunohistochemistry and flow cytometry, Malissen et al. demonstrated that high HVEM levels were associated with poor prognosis in patients with metastatic melanoma, as well as in other tumors, such as breast and gastric cancer [68, 71–74]. These results were validated by bioinformatic analysis of data from The Cancer Genome Atlas (TCGA) [75]. Multispectral immunofluorescence assays revealed that BTLA+ CD8+ T cells were physically related to HVEM+ tumor cells, thereby suggesting that HVEM molecules on melanoma cells inhibit infiltrating T cells via BTLA [75, 76]. This fully agrees with the results obtained in another independent study in melanoma patients, where it was observed that BTLA expression was reduced during differentiation of CD8+ T lymphocytes to effector cells, whereas *in vivo* expression was maintained in those lymphocytes specific for melanoma tumor antigens hindering their antitumor activity [77].

Despite BTLA-mediated inhibition of immune responses, this receptor can, promote positive signals through its Grb2 domain. Two studies on adoptive T-cell therapy in murine melanoma models described that BTLA- CD8+ T lymphocytes were not able to control tumor development *in vivo*, while their BTLA+ counterpart exhibited antitumor capacity, as well as greater survival and resistance to apoptosis [78, 79]. HVEM binding to BTLA induced inhibition of CD8+ T lymphocyte

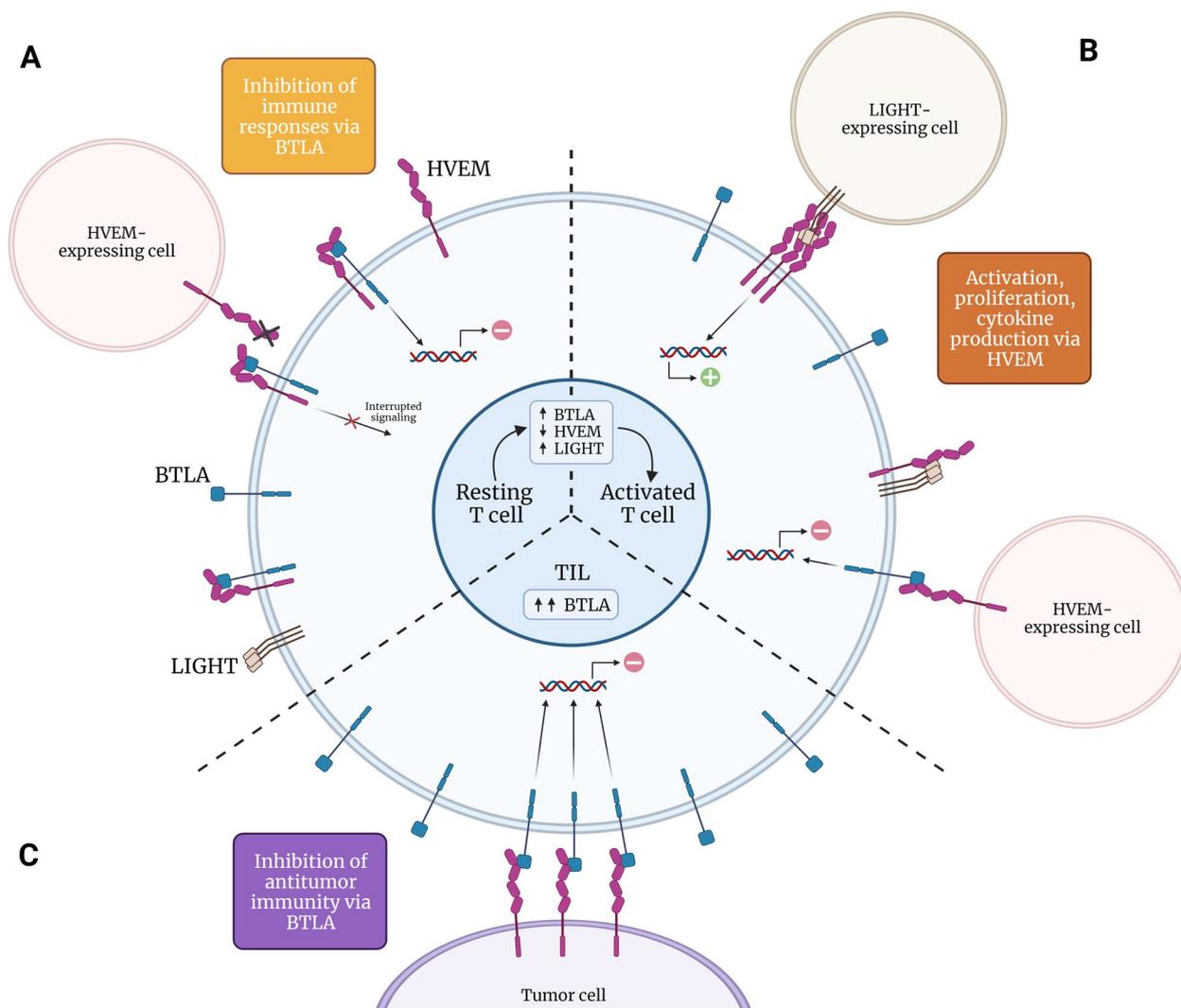


Fig. 3 BTLA-HVEM *cis* and *trans* signaling in different settings. **A** In resting T cells, BTLA inhibitory signaling plays a predominant role in *cis* interactions, impeding HVEM-mediated activation. **B** Upon activation, BTLA-HVEM *cis* complex is disrupted thus allowing *trans* interactions between BTLA and HVEM-expressing cells. **C** In the context of cancer, enhanced BTLA expression on tumor-infiltrating lymphocytes increase *trans* interaction and leads to the inhibition of T cell-mediated antitumor responses

proliferation as well as cytokine production, although AKT signaling was also triggered protecting T lymphocytes from apoptosis [79].

BTLA/HVEM axis in hematological cancers

Various studies have highlighted the importance of the BTLA/HVEM axis in tumor development, immunosuppression and prognosis. High HVEM and low BTLA mRNA levels in tumor cells from patients with follicular lymphoma (FL) and diffuse large B-cell lymphoma (DLBCL) were associated with shorter time-to-treatment, overall survival, and increased risk of transformation from FL to DLBCL [80, 81]. Contrarily, other works

support a tumor suppressor role for HVEM in these hematological tumors. Loss-of-function mutations are often detected in *TNFRSF14* gene that encodes this receptor, which are among the most frequently mutated genes in FL and DLBCL (up to 50% of cases), and associated with poor prognosis [80, 82–85]. Michael Boice et al. sustain that HVEM inactivation has a direct protumoral role in lymphoma cells, and also influences the TME. This hypothesis might have a simple explanation based on BTLA signaling. First, BTLA ITIM domains not only interact with SHP, but also with CD79, a typical BCR signaling mediator [86]. As a consequence, *trans* HVEM-BTLA interaction among adjacent tumor cells could lead

to BTLA-mediated transduction of negative signals to partially hinder tumor development, leading to a kind of contact inhibition and preventing tumor proliferation. Thus, disruption of the interaction between HVEM and BTLA through inactivating mutations or downregulation of these immune checkpoints provides a mechanism for stimulating BCR-related mitogenic signals in B-cell lymphoma cells [82]. This hypothesis about the tumor suppressive role of HVEM in lymphomas is supported by subsequent studies. Follicular helper T lymphocytes by decreasing BTLA surface expression, promoted the upregulation of the anti-apoptotic protein BCL-2 (via CD40L) in B germinal centers, favoring tumor growth [87, 88]. Therefore, according to these data, further studies are required to clarify more precisely the role played by the BTLA/HVEM axis in FL and DLBCL, either as a tumor suppressor or progression promoter by NF- κ B pathway activation.

Analysis of BTLA expression in 253 samples from patients with non-Hodgkin's lymphomas from germinal centers, including DLBCL, mantle cell lymphoma (MLC), marginal zone lymphoma, Burkitt's lymphoma and chronic lymphocytic leukemia (CLL) revealed that the latter displays the highest expression levels, suggesting that BTLA and HVEM expression on tumor cells from germinal centers could serve as a marker to distinguish between different types of non-Hodgkin's lymphomas [88]. CLL not only exhibits exacerbated BTLA levels, HVEM expression is also deeply dysregulated in this tumor. Moreover, higher *TNFRSF14* levels in leukemic cells were correlated with shorter overall survival [89]. In line with this, BTLA expression is significantly augmented in CD4+ and CD8+ T lymphocytes, as well as in NK cells. Importantly, increased levels of this inhibitory immune checkpoint on CD4+ T lymphocytes (but not CD8+) and NK cells correlated with shorter time to treatment in patients with CLL, thus suggesting an important role of BTLA dampening antitumor responses [89, 90]. In agreement, Alan G. Ramsay et al., analyzed the expression of BTLA in peripheral blood T lymphocytes from patients with CLL and found significantly higher BTLA levels compared to healthy donors [91].

Soluble form of BTLA in cancer

It is worth mentioning the existence of a soluble form of BTLA (sBTLA), as a consequence of alternative splicing [92]. Even though its role remains as of yet to be clarified, several studies have postulated this molecule as a prognostic factor or a predictor of treatment response. In solid tumors, elevated levels of sBTLA in sera have been associated with decreased survival and aggressive disease in prostate and ovarian cancer, pancreatic adenocarcinoma, clear cell renal cancer, and hepatocellular

carcinoma, among others [71, 93–96]. Circulating sBTLA levels have also been detected in patients with CLL [89, 97]. Indeed, increased sBTLA levels were associated with aggressive behavior and decreased time to treatment in these patients [89]. Noteworthy, further analyses of plasma sBTLA levels in patients with various solid tumors before undergoing ICB therapy brought to light that this molecule, beyond being an independent prognostic factor for survival or disease severity, may also serve as a valuable predictive tool for response to ICB-based therapy [98].

BTLA blockade as a target for cancer immunotherapy

Blockade of the inhibitory signaling pathway mediated by the BTLA/HVEM axis using antagonistic mAbs has been proposed as a potential therapeutic approach in multiple cancers. As an example, BTLA is overexpressed in CD8+ T lymphocytes specific for the tumor antigen NY-ESO-1 in melanoma. In fact, BTLA signaling disruption in vitro increased the proliferation and cytokine production in tumor-specific CD8+ T lymphocytes. Further, this approach showed a cooperative effect promoting T cell activity by blocking simultaneously other immune checkpoints, including TIM-3 and PD-1 [99]. Thus, combined BTLA and PD-1 blockade led to increased T lymphocyte activation, cytotoxicity, and cytokine production in response to allogeneic dendritic cells in vitro, unveiling the potential benefit of this combination [100]. Similar synergistic effects were achieved by combination with chemotherapy in ovarian carcinoma, where BTLA expression was related to poor outcome. BTLA blockade together with the chemotherapeutic agent paclitaxel significantly reduced tumor size and improved survival compared to both monotherapies in murine models of ovarian carcinoma. Furthermore, increased percentage of activated CD8+ and CD4+ T lymphocytes, higher levels of proinflammatory cytokines such as IL-12 or IFN- γ , as well as a reduction of various immunosuppressive cytokines (i.e. IL-10, IL-6 and TGF- β) were also observed by BTLA disruption [101].

Despite several studies strongly support the inhibitory and prognostic value of BTLA and HVEM in hematological cancers, there is scarce information to date about the antitumor effects associated to disruption of BTLA/HVEM axis. In-depth functional characterization is fundamental for safe implementation of BTLA blockade strategies into future clinical trials. As previously mentioned, BTLA expression is upregulated on the surface of CD4+ and CD8+ T lymphocytes as well as NK cells from patients with CLL, and negatively impacted patient's outcome. Treatment with an anti-BTLA agonistic antibody decreased IL-2 and IFN- γ production by cytotoxic lymphocytes from CLL patients, whereas BTLA blockade

restored cytokine production. Moreover, anti-BTLA blocking mAb promoted NK and T cell-mediated cytotoxicity, antibody-dependent cytotoxicity (ADCC) and was able to effectively eliminate leukemic cells ex vivo [89, 90].

Since reduced expression or deleterious mutations of HVEM was found to promote lymphomagenesis and tumor development in patients with FL and DLBCL, this molecule has been proposed as a possible therapeutic target [82]. In this context, a soluble form of HVEM (sHVEM) might be effective to restore tumor suppression upon binding to BTLA on cancer cells. In agreement, CD19-targeting and sHVEM-producing CAR-T cells were more effective than those exclusively based on CD19-CAR-T in xenograft murine lymphoma models, since sHVEM released locally within the tumor avoided undesired systemic immunosuppressive effects [82].

Allogeneic hematopoietic stem cell transplantation (allo-SCT) is a common treatment (sometimes the only curative option) in multiple hematological cancers. This approach enhances antitumor responses mediated by

alloreactive T lymphocytes directed against minor histocompatibility antigens (mHAg) [102, 103]. Increased BTLA expression was observed in mHAg-specific CD8+ T cells compared with memory effector CD8+ T cells in patients with acute myeloid leukemia and multiple myeloma, as well as constitutive expression of HVEM in mHAg tumor cells. Ex vivo BTLA blockade increased proliferation, cytokine production and degranulation of mHAg-specific CD8+ T lymphocytes. On this basis, targeting BTLA inhibitory signaling poses as a promising strategy to enhance T cell reactivity during allo-SCT [18]. Therapeutic alternatives targeting BTLA/HVEM axis currently explored in cancer are summarized in Fig. 4.

Clinical development of BTLA-targeted therapeutics

In 2019, it was approved the world’s first-in-class anti-BTLA blocking mAb tifcemalimab (icatolimab, TAB004/JS004, Junshi Biosciences), hindering BTLA/HVEM interaction. Tifcemalimab/icatolimab is an IgG₄ mAb, currently under clinical development in several

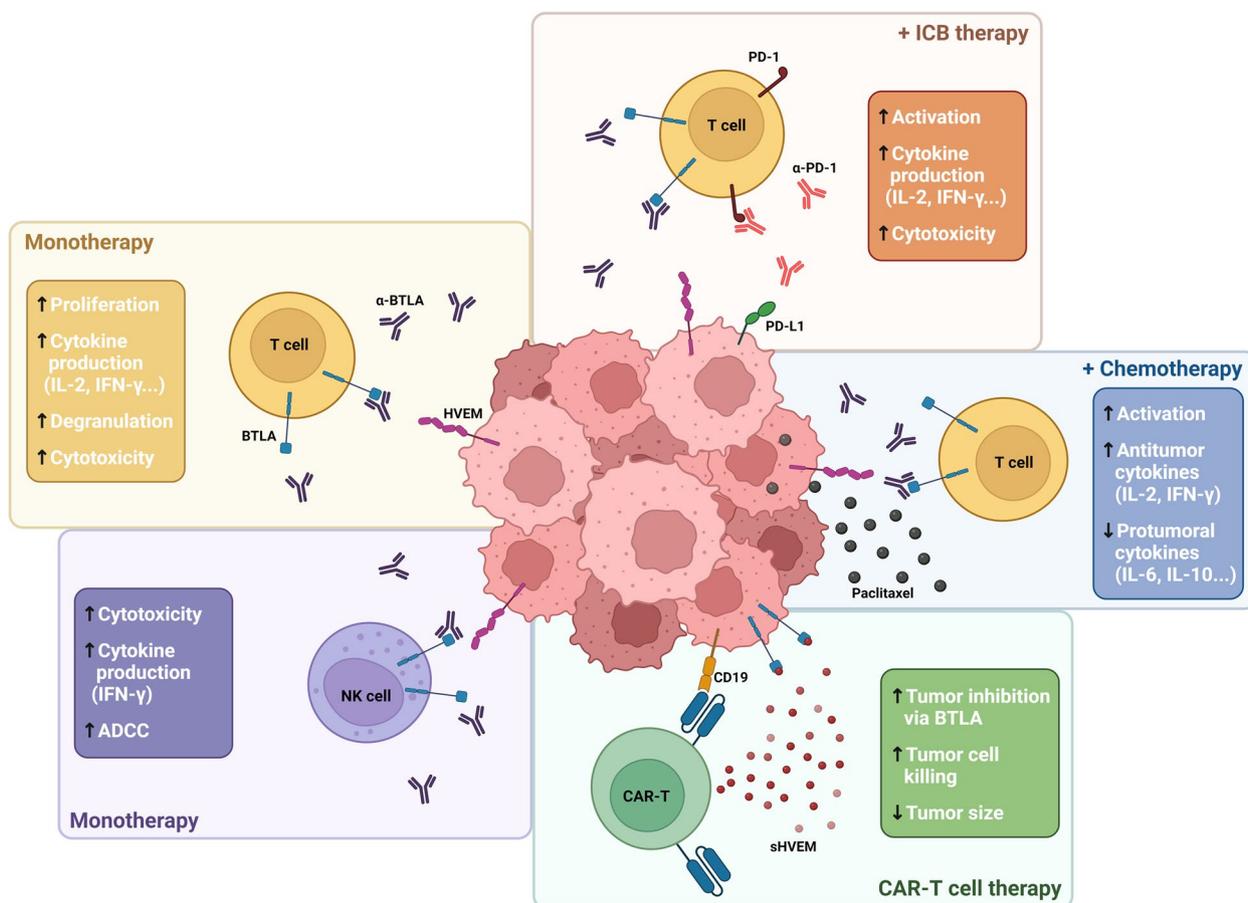


Fig. 4 BTLA/HVEM axis as a target for cancer immunotherapy. Multiple approaches are being developed at pre-clinical and clinical levels, including monoclonal antibodies (in monotherapy or combination with anti-PD-1 or chemotherapeutic agents) and sHVEM-producing CAR-T cells

solid tumors and hematological malignancies (summarized in Table 1). However, little data is available up to date regarding the clinical efficacy of this immunotherapeutic agent. In a phase Ia dose-escalation study (NCT04137900) with 25 patients with advanced solid tumors (median of 4 prior lines of therapy), including 15 patients who progressed upon anti-PD-1/PD-L1 therapy, tivecimalimab/icatolimab demonstrated a good safety profile and preliminary clinical efficacy (one partial response and six stable disease among 19 evaluable patients with a median follow-up of 32 weeks). Interestingly, joint expression of CD8 and HVEM has been proposed as a potential biomarker for positive response to BTLA blockade; however, there is scarce data available and extensive validation studies are required [104]. Forty-three patients with extensive-stage NSCLC refractory to prior therapies (including 14 patients who underwent anti-PD-1/PD-L1-based treatment) were enrolled in a phase I/II study (NCT05000684) exploring dual BTLA and PD-1 blockade. Despite preliminary data suggesting that tivecimalimab in combination with PD-1/PD-L1 axis blockade is well tolerated with 70.0% of the responses ongoing (median follow-up, 12.1 weeks), the median duration of response was not reached at the cut-off date [105].

Regarding hematological malignancies, a phase I study (NCT04477772) using tivecimalimab/icatolimab as monotherapy or in combination with the anti-PD-1 blocking mAb toripalimab is currently ongoing, with quite promising preliminary results. A total of 48 patients has been enrolled, including 20 non-Hodgkin's and 28

Hodgkin's lymphomas. All patients were heavily treated, with a median of 4 prior lines, including 32 patients who underwent anti-PD-1/PD-L1 therapy. With a median follow-up of 31.3 weeks, a total of one partial response (follicular lymphoma) and seven stable disease were observed in the tivecimalimab/icatolimab monotherapy cohort ($n=22$, including 11 individuals refractory to prior anti-PD-1/PD-L1 therapy). Among 12 evaluable patients with Hodgkin's lymphoma, one patient treated with dual BTLA/PD-1 blockade showed complete response, 4 of them partial response and 5 stable disease (including 7 individuals refractory to prior anti-CD30 therapy) [106, 107]. Interestingly, similar to solid tumors, a trend of correlation between high HVEM expression and clinical response was observed. Overall, treatment with tivecimalimab/icatolimab was well-tolerated and demonstrated preliminary clinical efficacy in solid tumors and lymphomas, thus encouraging further investigation of BTLA/HVEM axis as a promising target for immunotherapy.

Concluding remarks and future directions

ICB-based therapies have become an essential treatment option for a considerable number of tumors, despite the disappointing results obtained in others. On this basis, development of novel strategies for effectively targeting immune checkpoints represents a major milestone in cancer treatment to improve clinical efficacy and patient benefit by expanding the therapeutic spectrum.

As reviewed herein, recent accumulative evidence is shedding light on the key role of BTLA and HVEM in

Table 1 Current clinical trials targeting the BTLA/HVEM axis

Clinical trial	Disease	Therapy	Phase	Status
NCT04773951	Melanoma, renal carcinoma and urothelial carcinoma	Monotherapy and tivecimalimab/icatolimab + toripalimab (anti-PD-1)	Phase I	Recruiting
NCT05000684	Advanced lung cancer	Monotherapy and tivecimalimab/icatolimab + toripalimab (anti-PD-1)	Phase I	Recruiting
NCT05427396	Liver cancer, esophageal squamous cell carcinoma, gastric adenocarcinoma, cervical cancer, colorectal cancer	Tivecimalimab/icatolimab + toripalimab (anti-PD-1)	Phase I	Recruiting
NCT04929080	Head and neck cancer and nasopharyngeal carcinoma	Monotherapy and tivecimalimab/icatolimab + JS001 (anti-PD-1)	Phase I/II	Recruiting
NCT04477772	Recurrent/Refractory malignant lymphoma	Monotherapy and tivecimalimab/icatolimab + JS001 (anti-PD-1)	Phase I	Recruiting
NCT04137900	Advanced unresectable solid tumor and metastatic solid tumor	Tivecimalimab/icatolimab + toripalimab (anti-PD-1)	Phase I	Recruiting
NCT05664971	Advanced lung cancer	Tivecimalimab/icatolimab + JS001 (anti-PD-1) alone or in combination with chemotherapy	Phase I/II	Recruiting
NCT05891080	Stage III resectable or potentially resectable non-small cell lung cancer	Tivecimalimab/icatolimab + toripalimab (anti-PD-1) + chemotherapy + surgery	Phase II	Not recruiting yet
NCT05789069	Advanced renal cell carcinoma, melanoma, non-small cell lung cancer, gastric cancer, and colorectal cancer	HFB200603 (anti-BTLA) alone or in combination with tislelizumab (anti-PD-1)	Phase 1a/1b	Recruiting

tumor immune escape. In good agreement, dysregulation of BTLA/HVEM axis has been correlated with prognosis in both solid and hematological cancers. Moreover, circulating sBTLA has been revealed as a blood-based predictive biomarker of time to treatment and response to ICB-based therapies in various malignancies. Hence, BTLA/HVEM axis emerges as a novel and promising target for cancer immunotherapy. Accordingly, mounting preclinical data strongly support the function of BTLA dampening T lymphocyte and NK cell-mediated anti-tumor responses, which could be restored by employing blocking antibodies thereby subsequently triggering increased proliferation, cytokine production and cytotoxicity. As a next step into clinical application, the anti-BTLA blocking mAb tificemalimab/icatolimab has entered into various ongoing phase I/II clinical trials with very encouraging results so far. Thus, tificemalimab/icatolimab has demonstrated preliminary efficacy and safety profile in both solid and hematological malignancies. It is worth mentioning that, along with preclinical evidence, combination of anti-BTLA and current anti-PD-1/PD-L1 therapies has demonstrated promising results, even in those patients refractory to previous ICB monotherapy.

Besides cancer immunotherapy, it is worth mentioning that BTLA/HVEM axis is also likely to emerge as a critical player in other diseases, including sepsis [92, 108–110], neuroinflammation [111], infectious [112–114] and autoimmune diseases [115–119]. Three clinical trials using agonistic anti-BTLA antibodies (LY3361237, Stanford University and Eli Lilly and Company and ANB032 from AnaptysBio) are currently undergoing in patients with systemic lupus erythematosus (NCT05123586), Sjogren's syndrome (NCT05781451), and severe atopic dermatitis (NCT05935085). Moreover, agonistic anti-BTLA antibodies have been proposed as a potential therapy to ameliorate GvHD resulting from allogeneic bone marrow transplantation or to prolong allograft survival in models of renal or cardiac transplantation [120–125]. Nevertheless, studies targeting BTLA and HVEM are currently under preclinical development. In the framework of infectious diseases, preclinical data indicate that ICB could pose a highly relevant option in tuberculosis, malaria and HIV or hepatitis B infection [126, 127]. Nonetheless, despite several ongoing clinical trials using ICB-based therapies, as of yet, none of them targets BTLA/HVEM axis [126, 128].

Nevertheless, it should be noted the complexity of HVEM signaling, which is part of an intricate network constituted by *cis* and *trans* interactions with distinct binding partners. As a consequence, it appears reasonably essential to deepen understanding of the

functional interactome landscape of BTLA/HVEM axis in different cancer settings in order to establish individual therapeutic approaches for adequate patient stratification and safe clinical testing and to ultimately achieve the best outcomes for cancer patients.

In conclusion, compelling evidences indicate that BTLA/HVEM axis holds immense potential for future development of cancer immunotherapy strategies. As ongoing research are shedding new light on the complexity and intricate signaling interactions of BTLA/HVEM axis, therefore it poses as major challenges the identification of reliable predictive biomarkers to guide/monitor treatment response, and to delve into the optimization of combination therapies (mainly anti-PD-1/PD-L1-based therapies). Beyond the oncology field, the implication of BTLA/HVEM axis in transplantation, infections and autoimmune diseases offers promising avenues for investigation and, more importantly, opens up wide and great perspectives for future clinical application.

Acknowledgements

Not applicable.

Consent to publication

All authors agree to publish the present work.

Authors' contributions

Conceptualization, C.S-B; Writing—Original Draft Preparation, C.S.-B., S.L.-H., R.G.-D. and S.G.; Writing—Review and Editing, A.M.-P., C.A.-G., J.P.R. and J.M.G.-P. All authors have read and agreed to the published version of the manuscript.

Funding

This study was supported by the Spanish grant of Instituto de Salud Carlos III (PI22/00167 and CIBERONC CB16/12/00390), the Instituto de Investigación Sanitaria del Principado de Asturias (ISPA) (2021–079-INTRAMUR GALBAN-GOROS), Ayudas a Grupos PCTI Principado de Asturias (IDI/2021/000079), the Fundación Bancaria Cajastur, and the FEDER Funding Program from the European Union. C.S-B holds an Intramural ISPA-Janssen grant.

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

All authors declare no competing interests.

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Received: 7 June 2023 Accepted: 17 August 2023
Published online: 30 August 2023

References

- Sharma P, Siddiqui BA, Anandhan S, Yadav SS, Subudhi SK, Gao J, et al. The next decade of immune checkpoint therapy. *Cancer Discov*. 2021;11(4):838–57.
- Wolchok JD. Checkpoint blockade: the end of the beginning. *Nat Rev Immunol*. 2021;21(10):621.
- Pilard C, Ancion M, Delvenne P, Jerusalem G, Hubert P, Herfs M. Cancer immunotherapy: it's time to better predict patients' response. *Br J Cancer*. 2021;125(7):927–38.
- Amaria RN, Postow M, Burton EM, Tetzlaff MT, Ross MI, Torres-Cabala C, et al. Neoadjuvant relatlimab and nivolumab in resectable melanoma. *Nature*. 2022;611(7934):155–60.
- Mandel I, Haves Ziv D, Goldshtein I, Peretz T, Alishekevitz D, Fridman Dror A, et al. BND-22, a first-in-class humanized ILT2-blocking antibody, promotes antitumor immunity and tumor regression. *J Immunother Cancer*. 2022;10(9):e004859.
- Tawbi HA, Schadendorf D, Lipson EJ, Ascierto PA, Matamala L, Castillo Gutierrez E, et al. Relatlimab and nivolumab versus nivolumab in untreated advanced melanoma. *N Engl J Med*. 2022;386(1):24–34.
- Qin S, Xu L, Yi M, Yu S, Wu K, Luo S. Novel immune checkpoint targets: moving beyond PD-1 and CTLA-4. *Mol Cancer*. 2019;18(1):155.
- van Hall T, Andre P, Horowitz A, Ruan DF, Borst L, Zerbib R, et al. Monalizumab: inhibiting the novel immune checkpoint NKG2A. *J Immunother Cancer*. 2019;7(1):263.
- Battin C, Leitner J, Waidhofer-Sollner P, Grabmeier-Pfistershammer K, Olive D, Steinberger P. BTLA inhibition has a dominant role in the cis-complex of BTLA and HVEM. *Front Immunol*. 2022;13:956694.
- Sedy JR, Gavrieli M, Potter KG, Hurchla MA, Lindsley RC, Hildner K, et al. B and T lymphocyte attenuator regulates T cell activation through interaction with herpesvirus entry mediator. *Nat Immunol*. 2005;6(1):90–8.
- Hurchla MA, Sedy JR, Gavrieli M, Drake CG, Murphy TL, Murphy KM. B and T lymphocyte attenuator exhibits structural and expression polymorphisms and is highly induced in anergic CD4+ T cells. *J Immunol*. 2005;174(6):3377–85.
- Han P, Goularte OD, Rufner K, Wilkinson B, Kaye J. An inhibitory Ig superfamily protein expressed by lymphocytes and APCs is also an early marker of thymocyte positive selection. *J Immunol*. 2004;172(10):5931–9.
- Ward-Kavanagh LK, Lin WW, Sedy JR, Ware CF. The TNF Receptor superfamily in Co-stimulating and Co-inhibitory Responses. *Immunity*. 2016;44(5):1005–19.
- Steinberg MW, Cheung TC, Ware CF. The signaling networks of the herpesvirus entry mediator (TNFRSF14) in immune regulation. *Immunol Rev*. 2011;244(1):169–87.
- Gavrieli M, Murphy KM. Association of Grb-2 and PI3K p85 with phosphotyrosine peptides derived from BTLA. *Biochem Biophys Res Commun*. 2006;345(4):1440–5.
- Sedy JR, Ramezani-Rad P. HVEM network signaling in cancer. *Adv Cancer Res*. 2019;142:145–86.
- Zhao Q, Huang ZL, He M, Gao Z, Kuang DM. BTLA identifies dysfunctional PD-1-expressing CD4(+) T cells in human hepatocellular carcinoma. *Oncotarget*. 2016;5(12):e1254855.
- Hobo W, Norde WJ, Schaap N, Fredrix H, Maas F, Schellens K, et al. B and T lymphocyte attenuator mediates inhibition of tumor-reactive CD8+ T cells in patients after allogeneic stem cell transplantation. *J Immunol*. 2012;189(1):39–49.
- Oya Y, Watanabe N, Owada T, Oki M, Hirose K, Suto A, et al. Development of autoimmune hepatitis-like disease and production of autoantibodies to nuclear antigens in mice lacking B and T lymphocyte attenuator. *Arthritis Rheum*. 2008;50(8):2498–510.
- Watanabe N, Gavrieli M, Sedy JR, Yang J, Fallarino F, Loftin SK, et al. BTLA is a lymphocyte inhibitory receptor with similarities to CTLA-4 and PD-1. *Nat Immunol*. 2003;4(7):670–9.
- Krieg C, Boyman O, Fu YX, Kaye J. B and T lymphocyte attenuator regulates CD8+ T cell-intrinsic homeostasis and memory cell generation. *Nat Immunol*. 2007;8(2):162–71.
- Krieg C, Han P, Stone R, Goularte OD, Kaye J. Functional analysis of B and T lymphocyte attenuator engagement on CD4+ and CD8+ T cells. *J Immunol*. 2005;175(10):6420–7.
- Harrop JA, McDonnell PC, Brigham-Burke M, Lyn SD, Minton J, Tan KB, et al. Herpesvirus entry mediator ligand (HVEM-L), a novel ligand for HVEM/TR2, stimulates proliferation of T cells and inhibits HT29 cell growth. *J Biol Chem*. 1998;273(42):27548–56.
- Hurchla MA, Sedy JR, Murphy KM. Unexpected role of B and T lymphocyte attenuator in sustaining cell survival during chronic allostimulation. *J Immunol*. 2007;178(10):6073–82.
- Cheung TC, Steinberg MW, Osborne LM, Macauley MG, Fukuyama S, Sanjo H, et al. Unconventional ligand activation of herpesvirus entry mediator signals cell survival. *Proc Natl Acad Sci U S A*. 2009;106(15):6244–9.
- Sedy JR, Bjordahl RL, Bekiaris V, Macauley MG, Ware BC, Norris PS, et al. CD160 activation by herpesvirus entry mediator augments inflammatory cytokine production and cytolytic function by NK cells. *J Immunol*. 2013;191(2):828–36.
- Whitbeck JC, Peng C, Lou H, Xu R, Willis SH, de Ponce Leon M, et al. Glycoprotein D of herpes simplex virus (HSV) binds directly to HVEM, a member of the tumor necrosis factor receptor superfamily and a mediator of HSV entry. *J Virol*. 1997;71(8):6083–93.
- Ware CF, Sedy JR. TNF Superfamily Networks: bidirectional and interference pathways of the herpesvirus entry mediator (TNFRSF14). *Curr Opin Immunol*. 2011;23(5):627–31.
- Zhu Y, Yao S, Augustine MM, Xu H, Wang J, Sun J, et al. Neuron-specific SALM5 limits inflammation in the CNS via its interaction with HVEM. *Sci Adv*. 2016;2(4):e1500637.
- Cheung TC, Osborne LM, Steinberg MW, Macauley MG, Fukuyama S, Sanjo H, et al. T cell intrinsic heterodimeric complexes between HVEM and BTLA determine receptivity to the surrounding microenvironment. *J Immunol*. 2009;183(11):7286–96.
- Gonzalez LC, Loyet KM, Calemine-Fenaux J, Chauhan V, Wranik B, Ouyang W, et al. A coreceptor interaction between the CD28 and TNF receptor family members B and T lymphocyte attenuator and herpesvirus entry mediator. *Proc Natl Acad Sci U S A*. 2005;102(4):1116–21.
- Rooney IA, Butrovich KD, Glass AA, Borboroglu S, Benedict CA, Whitbeck JC, et al. The lymphotoxin-beta receptor is necessary and sufficient for LIGHT-mediated apoptosis of tumor cells. *J Biol Chem*. 2000;275(19):14307–15.
- Bechill J, Muller WJ. Herpesvirus entry mediator (HVEM) attenuates signals mediated by the lymphotoxin beta receptor (LTbetaR) in human cells stimulated by the shared ligand LIGHT. *Mol Immunol*. 2014;62(1):96–103.
- Murphy TL, Murphy KM. Slow down and survive: Enigmatic immunoregulation by BTLA and HVEM. *Annu Rev Immunol*. 2010;28:389–411.
- Steinberg MW, Huang Y, Wang-Zhu Y, Ware CF, Cheroutre H, Kronenberg M. BTLA interaction with HVEM expressed on CD8(+) T cells promotes survival and memory generation in response to a bacterial infection. *PLoS One*. 2013;8(10):e77992.
- Flynn R, Hutchinson T, Murphy KM, Ware CF, Croft M, Salek-Ardakani S. CD8 T cell memory to a viral pathogen requires trans costsignaling between HVEM and BTLA. *PLoS One*. 2013;8(10):e77991.
- Soroosh P, Doherty TA, So T, Mehta AK, Khorram N, Norris PS, et al. Herpesvirus entry mediator (TNFRSF14) regulates the persistence of T helper memory cell populations. *J Exp Med*. 2011;208(4):797–809.
- Crowe PD, VanArsdale TL, Walter BN, Ware CF, Hession C, Ehrensfels B, et al. A lymphotoxin-beta-specific receptor. *Science*. 1994;264(5159):707–10.
- Browning JL, Ngam-ek A, Lawton P, DeMarinis J, Tizard R, Chow EP, et al. Lymphotoxin beta, a novel member of the TNF family that forms a heteromeric complex with lymphotoxin on the cell surface. *Cell*. 1993;72(6):847–56.
- del Rio ML, Lucas CL, Buhler L, Rayat G, Rodriguez-Barbosa JI. HVEM/LIGHT/BTLA/CD160 costsignaling pathways as targets for immune regulation. *J Leukoc Biol*. 2010;87(2):223–35.
- Morel Y, de Schiano Colella JM, Harrop J, Deen KC, Holmes SD, Wattam TA, et al. Reciprocal expression of the TNF family receptor herpes virus

- entry mediator and its ligand LIGHT on activated T cells: LIGHT down-regulates its own receptor. *J Immunol.* 2000;165(8):4397–404.
42. Tamada K, Shimozaki K, Chapoval AI, Zhai Y, Su J, Chen SF, et al. LIGHT, a TNF-like molecule, costimulates T cell proliferation and is required for dendritic cell-mediated allogeneic T cell response. *J Immunol.* 2000;164(8):4105–10.
 43. Granger SW, Rickert S. LIGHT-HVEM signaling and the regulation of T cell-mediated immunity. *Cytokine Growth Factor Rev.* 2003;14(3–4):289–96.
 44. Yu KY, Kwon B, Ni J, Zhai Y, Ebner R, Kwon BS. A newly identified member of tumor necrosis factor receptor superfamily (TR6) suppresses LIGHT-mediated apoptosis. *J Biol Chem.* 1999;274(20):13733–6.
 45. Mauri DN, Ebner R, Montgomery RI, Kochel KD, Cheung TC, Yu GL, et al. LIGHT, a new member of the TNF superfamily, and lymphotoxin alpha are ligands for herpesvirus entry mediator. *Immunity.* 1998;8(1):21–30.
 46. Morel Y, Truneh A, Sweet RW, Olive D, Costello RT. The TNF superfamily members LIGHT and CD154 (CD40 ligand) costimulate induction of dendritic cell maturation and elicit specific CTL activity. *J Immunol.* 2001;167(5):2479–86.
 47. Jones A, Bourque J, Kuehm L, Opejin A, Teague RM, Gross C, et al. Immunomodulatory functions of BTLA and HVEM govern induction of extrathymic regulatory T cells and tolerance by dendritic cells. *Immunity.* 2016;45(5):1066–77.
 48. Le Bouteiller P, Barakonyi A, Giustiniani J, Lenfant F, Marie-Cardine A, Aguerre-Girr M, et al. Engagement of CD160 receptor by HLA-C is a triggering mechanism used by circulating natural killer (NK) cells to mediate cytotoxicity. *Proc Natl Acad Sci U S A.* 2002;99(26):16963–8.
 49. Agrawal S, Marquet J, Freeman GJ, Tawab A, Bouteiller PL, Roth P, et al. Cutting edge: MHC class I triggering by a novel cell surface ligand costimulates proliferation of activated human T cells. *J Immunol.* 1999;162(3):1223–6.
 50. Giustiniani J, Bensussan A, Marie-Cardine A. Identification and characterization of a transmembrane isoform of CD160 (CD160-TM), a unique activating receptor selectively expressed upon human NK cell activation. *J Immunol.* 2009;182(1):63–71.
 51. Cai G, Anumanthan A, Brown JA, Greenfield EA, Zhu B, Freeman GJ. CD160 inhibits activation of human CD4+ T cells through interaction with herpesvirus entry mediator. *Nat Immunol.* 2008;9(2):176–85.
 52. Abecassis S, Giustiniani J, Meyer N, Schiavon V, Ortonne N, Campillo JA, et al. Identification of a novel CD160+ CD4+ T-lymphocyte subset in the skin: a possible role for CD160 in skin inflammation. *J Invest Dermatol.* 2007;127(5):1161–6.
 53. Maeda M, Carpenito C, Russell RC, Dasanjh J, Veinotte LL, Ohta H, et al. Murine CD160, Ig-like receptor on NK cells and NKT cells, recognizes classical and nonclassical MHC class I and regulates NK cell activation. *J Immunol.* 2005;175(7):4426–32.
 54. Maiza H, Leca G, Mansur IG, Schiavon V, Boumsell L, Bensussan A. A novel 80-kD cell surface structure identifies human circulating lymphocytes with natural killer activity. *J Exp Med.* 1993;178(3):1121–6.
 55. Barakonyi A, Rabot M, Marie-Cardine A, Aguerre-Girr M, Polgar B, Schiavon V, et al. Cutting edge: engagement of CD160 by its HLA-C physiological ligand triggers a unique cytokine profile secretion in the cytotoxic peripheral blood NK cell subset. *J Immunol.* 2004;173(9):5349–54.
 56. Nikolova M, Marie-Cardine A, Boumsell L, Bensussan A. BY55/CD160 acts as a co-receptor in TCR signal transduction of a human circulating cytotoxic effector T lymphocyte subset lacking CD28 expression. *Int Immunol.* 2002;14(5):445–51.
 57. Giustiniani J, Marie-Cardine A, Bensussan A. A soluble form of the MHC class I-specific CD160 receptor is released from human activated NK lymphocytes and inhibits cell-mediated cytotoxicity. *J Immunol.* 2007;178(3):1293–300.
 58. Pasero C, Olive D. Interfering with coinhibitory molecules: BTLA/HVEM as new targets to enhance anti-tumor immunity. *Immunol Lett.* 2013;151(1–2):71–5.
 59. Pasero C, Speiser DE, Derre L, Olive D. The HVEM network: new directions in targeting novel costimulatory/co-inhibitory molecules for cancer therapy. *Curr Opin Pharmacol.* 2012;12(4):478–85.
 60. Shui JW, Steinberg MW, Kronenberg M. Regulation of inflammation, autoimmunity, and infection immunity by HVEM-BTLA signaling. *J Leukoc Biol.* 2011;89(4):517–23.
 61. Ware CF. Targeting the LIGHT-HVEM pathway. *Adv Exp Med Biol.* 2009;647:146–55.
 62. Ware CF. Targeting lymphocyte activation through the lymphotoxin and LIGHT pathways. *Immunol Rev.* 2008;223:186–201.
 63. Grabmeier-Pfistershammer K, Stecher C, Zettl M, Roskopf S, Rieger A, Zlabinger GJ, et al. Antibodies targeting BTLA or TIM-3 enhance HIV-1 specific T cell responses in combination with PD-1 blockade. *Clin Immunol.* 2017;183:167–73.
 64. Kubli SP, Berger T, Araujo DV, Siu LL, Mak TW. Beyond immune checkpoint blockade: emerging immunological strategies. *Nat Rev Drug Discov.* 2021;20(12):899–919.
 65. Oguro S, Ino Y, Shimada K, Hatanaka Y, Matsuno Y, Esaki M, et al. Clinical significance of tumor-infiltrating immune cells focusing on BTLA and Cbl-b in patients with gallbladder cancer. *Cancer Sci.* 2015;106(12):1750–60.
 66. Li X, Xu Z, Cui G, Yu L, Zhang X. BTLA expression in stage I–III non-small-cell lung cancer and its correlation with PD-1/PD-L1 and clinical outcomes. *Oncotargets Ther.* 2020;13:215–24.
 67. Ren S, Tian Q, Amar N, Yu H, Rivard CJ, Caldwell C, et al. The immune checkpoint, HVEM may contribute to immune escape in non-small cell lung cancer lacking PD-L1 expression. *Lung Cancer.* 2018;125:115–20.
 68. Lan X, Li S, Gao H, Nanding A, Quan L, Yang C, et al. Increased BTLA and HVEM in gastric cancer are associated with progression and poor prognosis. *Oncotargets Ther.* 2017;10:919–26.
 69. Kamal AM, Wasfey EF, Elghamry WR, Sabry OM, Elghobary HA, Radwan SM. Genetic signature of CTLA-4, BTLA, TIM-3 and LAG-3 molecular expression in colorectal cancer patients: Implications in diagnosis and survival outcomes. *Clin Biochem.* 2021;96:13–8.
 70. Li S, Dong C, Chen J, Gao X, Xie X, Zhang X. Identification of an immune checkpoint gene signature that accurately predicts prognosis and immunotherapy response in endometrial carcinoma. *Aging (Albany NY).* 2021;13(12):16696–712.
 71. Wang Q, Ye Y, Yu H, Lin SH, Tu H, Liang D, et al. Immune checkpoint-related serum proteins and genetic variants predict outcomes of localized prostate cancer, a cohort study. *Cancer Immunol Immunother.* 2021;70(3):701–12.
 72. Tsang JYS, Chan KW, Ni YB, Hlaing T, Hu J, Chan SK, et al. Expression and clinical significance of herpes virus entry mediator (HVEM) in breast cancer. *Ann Surg Oncol.* 2017;24(13):4042–50.
 73. Inoue T, Sho M, Yasuda S, Nishiwada S, Nakamura S, Ueda T, et al. HVEM expression contributes to tumor progression and prognosis in human colorectal cancer. *Anticancer Res.* 2015;35(3):1361–7.
 74. Migita K, Sho M, Shimada K, Yasuda S, Yamato I, Takayama T, et al. Significant involvement of herpesvirus entry mediator in human esophageal squamous cell carcinoma. *Cancer.* 2014;120(6):808–17.
 75. Malissen N, Macagno N, Granjeaud S, Granier C, Moutardier V, Gaudy-Marqueste C, et al. HVEM has a broader expression than PD-L1 and constitutes a negative prognostic marker and potential treatment target for melanoma. *Oncoimmunology.* 2019;8(12):e1665976.
 76. Ballotti R, Cheli Y, Bertolotto C. The complex relationship between MITF and the immune system: a Melanoma ImmunoTherapy (response) Factor? *Mol Cancer.* 2020;19(1):170.
 77. Derre L, Rivals JP, Jandus C, Pastor S, Rimoldi D, Romero P, et al. BTLA mediates inhibition of human tumor-specific CD8+ T cells that can be partially reversed by vaccination. *J Clin Invest.* 2010;120(1):157–67.
 78. Ritthipichai K, Haymaker CL, Martinez M, Aschenbrenner A, Yi X, Zhang M, et al. Multifaceted role of BTLA in the control of CD8(+) T-cell fate after antigen encounter. *Clin Cancer Res.* 2017;23(20):6151–64.
 79. Haymaker CL, Wu RC, Ritthipichai K, Bernatchez C, Forget MA, Chen JQ, et al. BTLA marks a less-differentiated tumor-infiltrating lymphocyte subset in melanoma with enhanced survival properties. *Oncoimmunology.* 2015;4(8):e1014246.
 80. Mintz MA, Cyster JG. T follicular helper cells in germinal center B cell selection and lymphomagenesis. *Immunol Rev.* 2020;296(1):48–61.
 81. Carreras J, Lopez-Guillermo A, Kikuti YY, Itoh J, Masashi M, Ikoma H, et al. High TNFRSF14 and low BTLA are associated with poor prognosis in Follicular Lymphoma and in Diffuse Large B-cell Lymphoma transformation. *J Clin Exp Hematop.* 2019;59(1):1–16.
 82. Boice M, Salloum D, Mourcin F, Sanghvi V, Amin R, Oricchio E, et al. Loss of the HVEM tumor suppressor in lymphoma and restoration by modified CAR-T cells. *Cell.* 2016;167(2):405–18 e13.

83. Okosun J, Bodor C, Wang J, Araf S, Yang CY, Pan C, et al. Integrated genomic analysis identifies recurrent mutations and evolution patterns driving the initiation and progression of follicular lymphoma. *Nat Genet.* 2014;46(2):176–81.
84. Morin RD, Mendez-Lago M, Mungall AJ, Goya R, Mungall KL, Corbett RD, et al. Frequent mutation of histone-modifying genes in non-Hodgkin lymphoma. *Nature.* 2011;476(7360):298–303.
85. Cheung KJ, Johnson NA, Affleck JG, Severson T, Steidl C, Ben-Neriah S, et al. Acquired TNFRSF14 mutations in follicular lymphoma are associated with worse prognosis. *Cancer Res.* 2010;70(22):9166–74.
86. Vendel AC, Calemine-Fenaux J, Izrael-Tomasevic A, Chauhan V, Arnott D, Eaton DL. B and T lymphocyte attenuator regulates B cell receptor signaling by targeting Syk and BLNK. *J Immunol.* 2009;182(3):1509–17.
87. Mintz MA, Felce JH, Chou MY, Mayya V, Xu Y, Shui JW, et al. The HVEM-BTLA axis restrains T cell help to germinal center B cells and functions as a cell-extrinsic suppressor in lymphomagenesis. *Immunity.* 2019;51(2):310–23 e7.
88. M'Hidi H, Thibault ML, Chetaille B, Rey F, Bouadallah R, Nicollas R, et al. High expression of the inhibitory receptor BTLA in T-follicular helper cells and in B-cell small lymphocytic lymphoma/chronic lymphocytic leukemia. *Am J Clin Pathol.* 2009;132(4):589–96.
89. Sordo-Bahamonde C, Lorenzo-Herrero S, Gonzalez-Rodriguez AP, Payer A, Gonzalez-Garcia E, Lopez-Soto A, et al. BTLA/HVEM Axis induces NK cell immunosuppression and poor outcome in chronic lymphocytic leukemia. *Cancers (Basel).* 2021;13(8):1766.
90. Sordo-Bahamonde C, Lorenzo-Herrero S, Martinez-Perez A, Gonzalez-Rodriguez AP, Payer AR, Gonzalez-Garcia E, et al. BTLA dysregulation correlates with poor outcome and diminished T cell-mediated anti-tumor responses in chronic lymphocytic leukemia. *Cancer Immunol Immunother.* 2023;72(7):2529–39.
91. Ramsay AG, Clear AJ, Fatah R, Gribben JG. Multiple inhibitory ligands induce impaired T-cell immunologic synapse function in chronic lymphocytic leukemia that can be blocked with lenalidomide: establishing a reversible immune evasion mechanism in human cancer. *Blood.* 2012;120(7):1412–21.
92. Monaghan SF, Banerjee D, Chung CS, Lomas-Neira J, Cygan KJ, Rhine CL, et al. Changes in the process of alternative RNA splicing results in soluble B and T lymphocyte attenuator with biological and clinical implications in critical illness. *Mol Med.* 2018;24(1):32.
93. Fanale D, Corsini LR, Brando C, Cutaia S, Di Donna MC, Filorizzo C, et al. Can circulating PD-1, PD-L1, BTN3A1, pan-BTN3As, BTN2A1 and BTLA levels enhance prognostic power of CA125 in patients with advanced high-grade serous ovarian cancer? *Front Oncol.* 2022;12: 946319.
94. Dong MP, Enomoto M, Thuy LTT, Hai H, Hieu VN, Hoang DV, et al. Clinical significance of circulating soluble immune checkpoint proteins in sorafenib-treated patients with advanced hepatocellular carcinoma. *Sci Rep.* 2020;10(1):3392.
95. Wang Q, Zhang J, Tu H, Liang D, Chang DW, Ye Y, et al. Soluble immune checkpoint-related proteins as predictors of tumor recurrence, survival, and T cell phenotypes in clear cell renal cell carcinoma patients. *J Immunother Cancer.* 2019;7(1):334.
96. Bian B, Fanale D, Dusetti N, Roque J, Pastor S, Chretien AS, et al. Prognostic significance of circulating PD-1, PD-L1, pan-BTN3As, BTN3A1 and BTLA in patients with pancreatic adenocarcinoma. *Oncoimmunology.* 2019;8(4):e1561120.
97. Landeira-Vinuela A, Arias-Hidalgo C, Juanes-Velasco P, Alcoceba M, Navarro-Bailon A, Pedreira CE, et al. Unravelling soluble immune checkpoints in chronic lymphocytic leukemia: physiological immunomodulators or immune dysfunction. *Front Immunol.* 2022;13:965905.
98. Gorgulho J, Roderburg C, Heymann F, Schulze-Hagen M, Beier F, Vucur M, et al. Serum levels of soluble B and T lymphocyte attenuator predict overall survival in patients undergoing immune checkpoint inhibitor therapy for solid malignancies. *Int J Cancer.* 2021;149(5):1189–98.
99. Fourcade J, Sun Z, Pagliano O, Guillaume P, Luescher IF, Sander C, et al. CD8(+) T cells specific for tumor antigens can be rendered dysfunctional by the tumor microenvironment through upregulation of the inhibitory receptors BTLA and PD-1. *Cancer Res.* 2012;72(4):887–96.
100. Stecher C, Battin C, Leitner J, Zettl M, Grabmeier-Pfistershammer K, Holler C, et al. PD-1 blockade promotes emerging checkpoint inhibitors in enhancing T cell responses to allogeneic dendritic cells. *Front Immunol.* 2017;8:572.
101. Chen YL, Lin HW, Chien CL, Lai YL, Sun WZ, Chen CA, et al. BTLA blockade enhances Cancer therapy by inhibiting IL-6/IL-10-induced CD19(high) B lymphocytes. *J Immunother Cancer.* 2019;7(1):313.
102. Bleakley M, Riddell SR. Molecules and mechanisms of the graft-versus-leukaemia effect. *Nat Rev Cancer.* 2004;4(5):371–80.
103. Goulmy E. Minor histocompatibility antigens: allo target molecules for tumor-specific immunotherapy. *Cancer J.* 2004;10(1):1–7.
104. Schilder RJ, Powderly JD, Park H, Bilen MA, McKean M, May R, et al. Phase Ia dose-escalation study of the anti-BTLA antibody icatolimab as a monotherapy in patients with advanced solid tumor. 2022;40(16_suppl):2643. https://ascopubs.org/doi/abs/10.1200/JCO.2022.40.16_suppl.2643.
105. Wang J, Yu Y, Wang Q, Yang R, Lv D, Xia B, et al. Phase I/II combination study of tificemalimab with toripalimab in patients with refractory extensive stage small cell lung cancer (ES-SCLC). 2023;41(16_suppl):8579. https://ascopubs.org/doi/abs/10.1200/JCO.2023.41.16_suppl.8579#:~:text=Conclusions%3A%20Tificemalimab%20in%20combination%20with,treatment%20in%20SCLC%20is%20warranted.
106. Ma J, Xie Y, Zhang H, Song Y, Zhao W, Pan Y, et al. Phase I study of the anti-BTLA antibody icatolimab as a single agent or in combination with toripalimab in relapsed/refractory lymphomas. 2022;40(16_suppl):7578. https://ascopubs.org/doi/abs/10.1200/JCO.2022.40.16_suppl.7578#:~:text=Conclusions%3A%20Icatolimab%20alone%20or%20in,patients%20with%20R%2FR%20lymphoma.
107. Ma J, Song Y, Xie Y, Zhang H-L, Shuang Y, Peng Z, et al. Phase I study of the anti-btla antibody tificemalimab as a single agent or in combination with toripalimab in relapsed/refractory lymphomas. *Blood.* 2022;140(Supplement 1):3716–7.
108. Shubin NJ, Chung CS, Heffernan DS, Irwin LR, Monaghan SF, Ayala A. BTLA expression contributes to septic morbidity and mortality by inducing innate inflammatory cell dysfunction. *J Leukoc Biol.* 2012;92(3):593–603.
109. Lange A, Sunden-Cullberg J, Magnuson A, Hultgren O. Soluble B and T lymphocyte attenuator correlates to disease severity in sepsis and high levels are associated with an increased risk of mortality. *PLoS One.* 2017;12(1):e0169176.
110. Wang WD, Yang XR, Guo MF, Pan ZF, Shang M, Qiu MJ, et al. Up-regulation of BTLA expression in myeloid dendritic cells associated with the treatment outcome of neonatal sepsis. *Mol Immunol.* 2021;134:129–40.
111. Joseph J, Premeaux TA, Pinto DO, Rao A, Guha S, Panfil AR, et al. Retroviral b-Zip protein (HBZ) contributes to the release of soluble and exosomal immune checkpoint molecules in the context of neuroinflammation. *J Extracell Biol.* 2023;2(7):e102.
112. Shen X, Zhang J, Tang P, Song H, Liu X, Huang Z, et al. Expression and clinical significance of B and T lymphocyte attenuator on CD4(+) and CD8(+) T cells from patients with pulmonary tuberculosis. *Indian J Pathol Microbiol.* 2019;62(2):232–8.
113. Serriari NE, Gondois-Rey F, Guillaume Y, Remmerswaal EB, Pastor S, Mesal N, et al. B and T lymphocyte attenuator is highly expressed on CMV-specific T cells during infection and regulates their function. *J Immunol.* 2010;185(6):3140–8.
114. Cai G, Nie X, Li L, Hu L, Wu B, Lin J, et al. B and T lymphocyte attenuator is highly expressed on intrahepatic T cells during chronic HBV infection and regulates their function. *J Gastroenterol.* 2013;48(12):1362–72.
115. Yang B, Huang Z, Feng W, Wei W, Zhang J, Liao Y, et al. The Expression of BTLA was increased and the expression of HVEM and LIGHT was decreased in the T cells of patients with rheumatoid arthritis [corrected]. *PLoS One.* 2016;11(5):e0155345.
116. Oster C, Wilde B, Specker C, Sun M, Kribben A, Witzke O, et al. BTLA expression on Th1, Th2 and Th17 effector t-cells of patients with systemic lupus erythematosus is associated with active disease. *Int J Mol Sci.* 2019;20(18):4505.
117. Inuo M, Ihara K, Matsuo T, Kohno H, Hara T. Association study between B- and T-lymphocyte attenuator gene and type 1 diabetes mellitus or systemic lupus erythematosus in the Japanese population. *Int J Immunogenet.* 2009;36(1):65–8.
118. Huarte E, Jun S, Rynda-Apple A, Golden S, Jackiw L, Hoffman C, et al. Regulatory T cell dysfunction acquiesces to BTLA+ regulatory B cells subsequent to oral intervention in experimental autoimmune encephalomyelitis. *J Immunol.* 2016;196(12):5036–46.

119. Wojciechowicz K, Spodzieja M, Lisowska KA, Wardowska A. The role of the BTLA-HVEM complex in the pathogenesis of autoimmune diseases. *Cell Immunol.* 2022;376:104532.
120. Zhang J, Zhang H, Wang Z, Yang H, Chen H, Cheng H, et al. BTLA suppress acute rejection via regulating TCR downstream signals and cytokines production in kidney transplantation and prolonged allografts survival. *Sci Rep.* 2019;9(1):12154.
121. Wang Z, Yang H, Liu X, Zhang J, Han Z, Tao J, et al. Role of B and T lymphocyte attenuator in renal transplant recipients with biopsy-proven acute rejection. *Med Sci Monit.* 2018;24:387–96.
122. Uchiyama M, Jin X, Matsuda H, Bashuda H, Imazuru T, Shimokawa T, et al. An agonistic anti-BTLA mAb (3C10) induced generation of IL-10-dependent regulatory CD4⁺ T cells and prolongation of murine cardiac allograft. *Transplantation.* 2014;97(3):301–9.
123. Sakoda Y, Park JJ, Zhao Y, Kuramasu A, Geng D, Liu Y, et al. Dichotomous regulation of GVHD through bidirectional functions of the BTLA-HVEM pathway. *Blood.* 2011;117(8):2506–14.
124. Albring JC, Sandau MM, Rapaport AS, Edelson BT, Satpathy A, Mashayekhi M, et al. Targeting of B and T lymphocyte associated (BTLA) prevents graft-versus-host disease without global immunosuppression. *J Exp Med.* 2010;207(12):2551–9.
125. Zhang H, Wang Z, Zhang J, Zhang X, Gui Z, Sun L, et al. The synergism of B and T lymphocyte attenuator (BTLA) and cytotoxic T lymphocyte associated antigen-4 (CTLA-4) attenuated acute T-cell mediated rejection and prolonged renal graft survival. *Transl Androl Urol.* 2020;9(5):1990–9.
126. Wykes MN, Lewin SR. Immune checkpoint blockade in infectious diseases. *Nat Rev Immunol.* 2018;18(2):91–104.
127. Gay CL, Bosch RJ, Ritz J, Hataye JM, Aga E, Tressler RL, et al. Clinical Trial of the Anti-PD-L1 Antibody BMS-936559 in HIV-1 Infected Participants on Suppressive Antiretroviral Therapy. *J Infect Dis.* 2017;215(11):1725–33.
128. Rasmussen TA, Anderson JL, Wightman F, Lewin SR. Cancer therapies in HIV cure research. *Curr Opin HIV AIDS.* 2017;12(1):96–104.

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